

L. monocytogenes Serovar 4a is a Possible Evolutionary Intermediate Between *L. monocytogenes* Serovars 1/2a and 4b and *L. monocytogenes* Serovar 4a

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The genetic relationships between serovars 1/2a, 4a, and 4b of *L. monocytogenes* were investigated by using pulsed-field gel electrophoresis (PFGE) and phylogenetic analysis. Serovar 4a was found to be a possible evolutionary intermediate between serovars 1/2a and 4b. The genetic relationships between serovars 1/2a, 4a, and 4b were confirmed by comparing the DNA profiles of 23S rRNA and 16S rRNA. The genetic relationships between serovars 1/2a, 4a, and 4b were confirmed by comparing the DNA profiles of 23S rRNA and 16S rRNA. The genetic relationships between serovars 1/2a, 4a, and 4b were confirmed by comparing the DNA profiles of 23S rRNA and 16S rRNA.

athgenic lipid I (LIPI-1) and eelital gene (i.e., A and B). This gene that a tlf, m h izital gene tafe, deletion, fke i lence-a i ciated gene in the genome of a commu a tice tl ma leadtl be etlchange in the ce i nding her tl e in cl ding athgenic [2, 5, 13].

Next, the h l genetic elati hi amug a i . e i a and . a e fa f m clea i the ba i f tl die i the i lence-a i ciated gene cha A, B, and ite ralt gene . T ga i r h l genetic i ght i tl the . g i e a l zed the r cle t l d e e rce f 23S RNA and 16S RNA and the gene cl tle 0029- 0042, B- , - C, and - i . e i a 1/2a, 4a, and 4b and . The at i rale tl fi c i the e gene egi i e ba ed i the e i i b e at i r that the 0029- 0042 gene egi i i l tl tle i e be i e e r . and [13], and that the P fA- eg lated i lence gene cl tle i e et i r . , , and . b tl ab et l i . , , and [11]. In additi r , e a e ed the tlal f . - ecific A and B, and 10 . - ecific gene i the e ecie / e i a , al r g i h her tl ic cha ct e izati r b i r g and i ced e . The e l ha ic a i ache ha e i e f i l i r e et l del i e at i r i f ecie and cha ct e izati r i f at lical . and . tl a r [18, 32, 38–40].

MATERIALS AND METHODS

Bacterial Strains

A total of 32 strains were examined in this study (Table 1). These included 25 strains/isolates, four of which came from reference collections, and 21 were isolated from food products and processing plants and vessels [41]. In addition, two (ATCC 33090 and AB2497), one (Li01), one (C15), one (ATCC 35967), and two (Li07 and Li08) strains were acquired from reference collections (Table 1). strains were refreshed from glycerol stocks maintained at -80°C and cultured on tryptic soy agar plates with 7% sheep blood, followed by growth in brain heart infusion broth (BHI; Oxoid, Hampshire, England) at 37°C.

Mouse Virulence Assay

The virulence potential of 25 and one (ATCC 33090) strains was assessed in accordance with a previously reported protocol [17]. Briefly, female ICR mice at 20–22 g (Zhejiang College of Traditional Chinese Medicine, Hangzhou, China) were allowed to acclimatize for 3 days. Five groups of mice (six per group) were inoculated intraperitoneally with 0.2-ml aliquots of appropriately diluted strain resuspended in phosphate-buffered saline (PBS, 0.01 M, pH 7.2). Mice in the control group were injected with 0.2 ml of PBS. The LD₅₀ values were calculated by using the trimmed Spearman-Karber method on the basis of mouse mortality data recorded

during a 10-day post-injection period, and the relative virulence (%) of these strains was determined as described previously [19].

Plaque-Forming Assay

The ability of strains to form plaques on mouse fibroblasts L929 cells was assessed as described previously [16]. Cell monolayers were grown to 80% confluence in 2 ml of DMEM containing 10% fetal bovine serum in 6-well plates (Corning, U.S.A.). The overnight cultures were centrifuged and resuspended in PBS. For each strain tested, one well was infected with 5×10⁵ CFU and the other was infected with 1.5×10⁵ CFU. Upon 1-h incubation at 37°C, the cell monolayers were washed three times with PBS and overlaid with 3 ml of DMEM containing 20 µg/ml gentamicin and 1.4% agarose (Oxoid Ltd., Hampshire, England). Following a 3-day incubation at 37°C, a second 2-ml overlay of DMEM containing 0.02% neutral red solution and 1.4% agarose was added. After a final day of incubation, plaques were photographed by a digital camera. The diameters of 25 plaques were measured using Adobe Photoshop software for each strain. The plaque size of reference strain 10403S was set at 100%.

Assays for Hemolytic and Phospholipase Activities

Hemolytic activity of strains was assayed in sheep blood agar plates as previously described [8]. To titrate the hemolytic activity, supernatant from BHI broth cultures was serially diluted by 2-fold in a 96-well V-bottom microplate with saline (8.5 g/l NaCl). An equal volume of sheep red blood cells in saline was added to each well and the microplates were incubated at 37°C for 1 h. The hemolytic titer of each strain is expressed as the reciprocal of the corresponding dilution of the supernatant required to lyse 50% of the erythrocytes in triplicate wells [16]. Phospholipase activity of strains was examined with the egg yolk assay of Ermolaeva [6] without charcoal activation. The BHI agar plates were supplemented with 5% fresh egg yolk suspension in saline. cultures were streaked onto the plates and incubated at 37°C for 48 h, with Li01 being applied as the positive control displaying an opacity zone surrounding the streak [9].

PCR

One ml of each broth culture was transferred to an Eppendorf tube and centrifuged at 12,000 × for 3 min. The cell pellet was washed twice with milli-Q water (Millipore China Ltd, Beijing, China) and then resuspended in TZ buffer (2% Triton X-100, 2.5 mg/ml NaN₃, and Tris-HCl, pH 8.0). After boiling for 10 min, the bacterial suspension was cooled on ice for 5 min and subsequently centrifuged at 12,000 × at 4°C for 1 min. The resulting supernatant was used as template DNA. The PCR mixture (in a volume of 30 µl) was made up of 3 µl of 10×PCR buffer [200 mM Tris-HCl, pH 9.0, 100 mM KCl, 20 mM MgCl₂, 100 mM (NH₄)₂SO₄, and 1% Triton X-100], 0.6 µl of dNTPs (10 mM), 0.6 µl of each primer (5 µM, custom synthesized by Invitrogen Biotechnology Co. Ltd., Shanghai, China), 0.8 µl of DNA polymerase (2 U/µl; TaKaRa Biotech Co. Ltd., Dalian, China), and milli-Q water to a final volume of 28 µl, and 2 µl template DNA. To amplify products larger than 4 kb, DNA polymerase (TaKaRa) was utilized. The reaction mixtures were subjected to a hot start at 95°C for 3 min prior to 25 cycles of amplification, with a final extension at 72°C for 5 min in a thermal cycler (MJ Research Inc., Boston, MA, U.S.A.). The annealing temperatures varied with specific primer pairs (Supplementary Table 1), and the duration of extension depended on the length of amplicons (1 min per kb, at 72°C). The PCR-

Table 1. Characteristics of *Listeria* strains used in this study.

Strain	Serovar	Source	Hemolytic titer	Relative size of plaque (%) ^a	Mouse mortality (dead/tested) ^b	Relative virulence ^c	logLD ₅₀ ^d
<i>L. monocytogenes</i> EGD	1/2a	Reference strain	2 ²	ND	11/30	36.6%	6.64
10403S	1/2a	Reference strain	2 ²	100.0	18/30	60%	5.49
NICPBP54006	4a	Reference strain	2 ²	0	1/30	3.3%	8.35
NICPBP54007	4b	Reference strain	2 ²	ND	11/30	36.6%	6.79
mLm3	4b	Raw milk	2 ³	108.3	28/30	93.3%	3.86
mLm4	4a	Pasteurized milk	2 ³	0	2/30	6.6%	8.14
mLm10	1/2a	Pasteurized milk	2 ²	95.7	18/30	60%	5.55
fLm1	1/2a	Beef	2 ²	96.3	14/30	46.6%	6.26
fLm2	1/2b	Pork chops	2 ²	88.8	13/30	43.3%	6.45
fLm3	1/2a	Raw pork	2 ²	98.3	15/30	50%	6.07
fLm4	1/2c	Vegetable	2 ²	85.0	15/30	50%	6.11
fLm5	1/2b	Chicken	2 ¹	92.0	16/30	53.3%	5.83
eLm1	1/2a	Seafood plant sewage	2 ³	103.7	18/30	60%	5.53
eLm2	1/2b	Milk plant vessel	2 ²	102.8	12/30	40%	6.46
eLm3	1/2b	Milk plant sewage	2 ²	83.7	12/30	40%	6.43
eLm4	1/2b	Milk plant sewage	2 ²	97.0	13/30	43.3%	6.32
eLm5	1/2a	Milk plant vessel	2 ²	89.3	18/30	60%	5.45
sLm1	4b	American red drum	2 ²	84.5	11/30	36.6%	6.74
sLm2	1/2c	American red drum	2 ¹	92.0	14/30	46.6%	6.19
sLm3	4b	American red drum	2 ²	85.6	11/30	36.6%	6.72
sLm4	1/2b	Shelled shrimps	2 ²	102.3	16/30	53.3%	5.94
sLm5	4b	Shelled shrimps	2 ²	90.1	25/30	83.3%	4.40
sLm6	1/2b	Shelled shrimps	2 ²	91.9	17/30	56.6%	5.79
sLm7	1/2b	Shelled shrimps	2 ²	100.4	21/30	70%	5.08
sLm8	1/2a	Shelled shrimps	2 ²	98.8	13/30	43.3%	6.31
<i>L. innocua</i> ATCC 33090	6a	Reference strain	<2 ⁰	0	0/30	0%	ND
AB2497	6a	Reference strain	<2 ⁰	ND	ND	ND	ND
<i>L. ivanovii</i> Li01	5	Reference strain	2 ⁴	ND	ND	ND	ND
<i>L. welshimeri</i> C15		Reference strain	<2 ⁰	ND	ND	ND	ND
<i>L. seeligeri</i> ATCC 35967		Reference strain	2 ¹	ND	ND	ND	ND
<i>L. grayi</i> Li07		Reference strain	<2 ⁰	ND	ND	ND	ND
Li08		Reference strain	<2 ⁰	ND	ND	ND	ND

^a Titer (CFU/ml) of 10403S, 100%.
^b Listeria (IC₅₀) of 10⁸, 10⁷, 10⁶, 10⁵, 10⁴, r, .
^c Relative virulence (%) of 10⁻, r, .ND,
^d Titer (LD₅₀) of -K r r, r, LD₅₀ r, r, .

amplified products were electrophoresed on 1.0% agarose gel in the presence of ethidium bromide (0.5 µg/ml) and visualized under UV transillumination. The cluster (and its equivalent in other strains) and three specific internalin gene clusters (, , and) were amplified with primers targeting their flanking genes (, / , / , and /). The full-length sequences of LIPI-1 between and were covered by five fragments in separate PCRs. In addition, primers were derived from -specific genes , , , , , , , and [10] for sequence comparison among species (Supplementary Table 1).

Cloning and Sequencing of PCR Products

PCR fragments were purified by using the AxyPrep DNA Gel Extraction Kit (Axygen Inc., U.S.A.) and inserted by T-A cloning strategy into the pMD18-T vector (TaKaRa). The recombinant plasmids

were introduced into DH5 and confirmed by PCR and restriction digestion with EcoRI and HindIII. The positive clones were selected and sequenced by the dideoxy method on an ABI-PRISM 377 DNA sequencer.

Genome Walking

Additional primers for genome walking were designed from the gene regions whose sequences became available in the study. Nested PCR was performed by using the TaRaKa Genome Walking Kit in accordance with the procedures recommended by the manufacturer.

Phylogenetic Analysis

Deduced amino acid sequences of the ORFs under investigation were aligned by ClustalX software (version 1.8). The corresponding nucleotide sequences were then trimmed and aligned [32]. Phylogenetic and molecular analyses were undertaken by using the Molecular Evolutionary Genetics Analysis software (MEGA version 3.0) (<http://>

(www.megasoftware.net). Phylogenetic trees were constructed and compared by using neighbor-joining (NJ), maximum parsimony (MP), minimum evolution (ME), and UPGMA methods [17, 36]. The robustness of the branching pattern was tested by bootstrap analyses through 1,000 replications.

GenBank Accession Numbers

Forty-five nucleotide sequences covering the genes of *L. monocytogenes* strains examined in this study have been deposited in GenBank (Accession Nos. EF392667 to EF392669, EF690661 to EF690672, EU073135 to EU073161, and EU444834 to EU444836) (Supplementary Table 2).

RESULTS

Virulence to Mice

When each of the 23 *L. monocytogenes* strains belonging to serovars 1/2a, 1/2b, 1/2c, and 4b was inoculated into ICR mice intraperitoneally at a dose of 10^8 CFU, the mortality rates were 11–28% of 30 mice tested, and the LD₅₀ values were 36.3%–93.3% (Table 1).

(3.86–6.79), the *L. monocytogenes* strains belonging to serovar 4a (NICPBP54006 and mLm4) exhibited minimal lethality (with mortality rates of 1 and 2% of 30 mice tested, and the LD₅₀ values were 3.3 and 6.6%, and $LD_{50} > 8$) (Table 1). The ATCC 33090 strain was highly lethal in mice (mortality rate of 100% and $LD_{50} = 1$) (Table 1).

Plaque-Forming Ability

The plaque-forming ability of *L. monocytogenes* strains in L929 cells was determined. The plaque-forming ability of serovars 1/2a, 1/2b, 1/2c, and 4b was measured. The plaque-forming ability of serovar 1/2a was 83.7%–108.3%, whereas that of serovar 4a was 100%. The plaque-forming ability of strains NICPBP54006 and mLm4 and ATCC 33090 was determined. The plaque-forming ability of ATCC 33090 was 100% in L929 cells (Table 1), which was significantly higher than that of the other strains.

Hemolytic and Phospholipase Activities

The hemolytic and phospholipase activities of serovar 4a strains (NICPBP54006 and mLm4) were determined. The hemolytic activity of these strains was

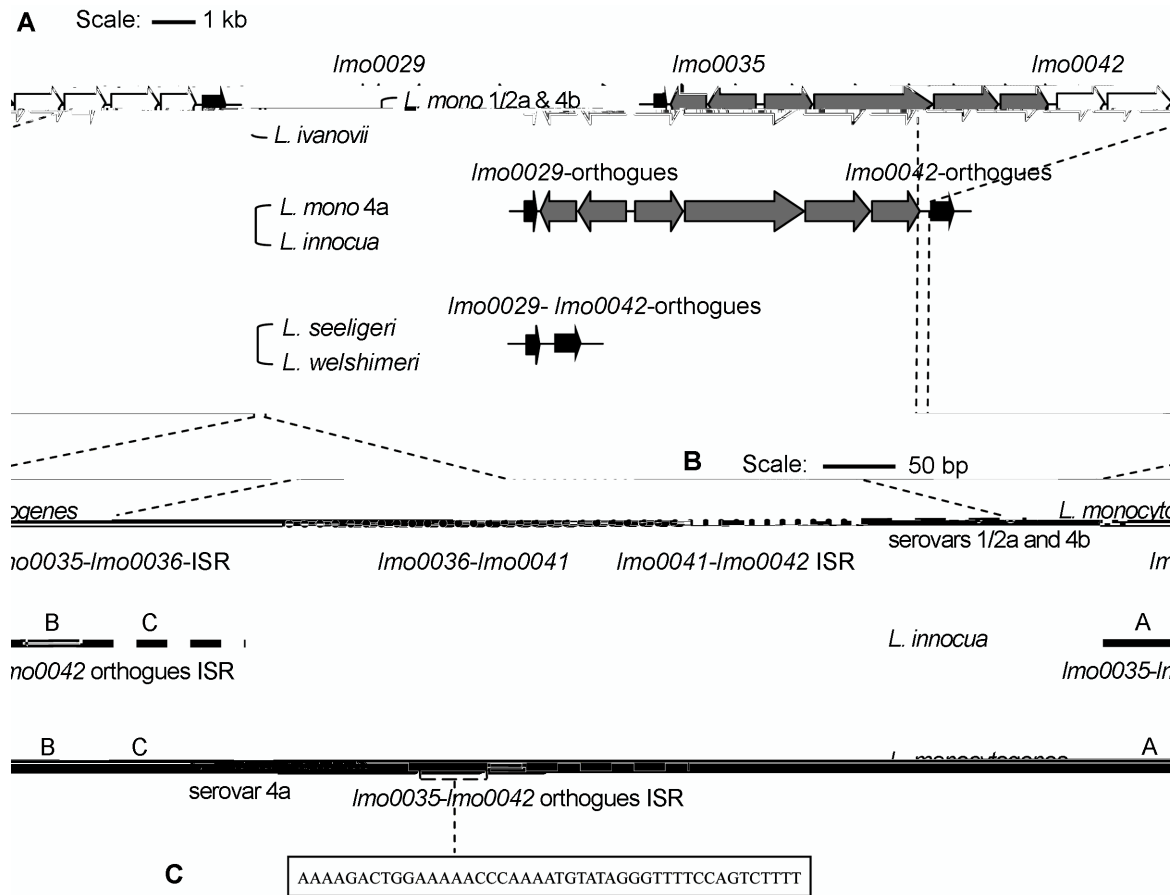


Fig. 1. A. Genetic structures of the *L. monocytogenes* *Imo0029-0042* region and its orthologs in *Listeria* species. B. Genetic organization of the *Imo0035-Imo0042* intergenic spacer region (ISR) of *L. monocytogenes* serovars 1/2a and 4b in relation to those of *L. monocytogenes* serovar 4a and *L. innocua*. The *Imo0035-Imo0042* ISR of *L. monocytogenes* serovar 4a contains three segments from different origins (see text for details). C. Alignment of segment A of the *Imo0035-Imo0042* ISR with putative insertion junctions.

Table 2. Comparison of nucleotide sequences in the *lmo0029-0042* locus among *Listeria* species.

Strain	Length (bp)	Nucleotide identity (%)			
		<i>L. monocytogenes</i>		<i>L. innocua</i>	<i>L. welshimeri</i>
		EGD (1/2a)	F2365 (4b)	CLIP11262	SLCC5334
<i>L. monocytogenes</i> 54007 (4b)	15,391	95.3	98.4	85.3	82.6
<i>L. monocytogenes</i> 54006 (4a)	8,735	87.8	88.5	89.2	78.0
<i>L. monocytogenes</i> mLm4 (4a)	8,735	87.6	88.4	89.1	78.1
<i>L. innocua</i> ATCC33090	8,735	85.9	85.8	99.6	77.5
<i>L. welshimeri</i> C15	1,189	82.6	82.2	76.8	98.6
<i>L. seeligeri</i> ATCC35967	1,189	80.8	80.1	72.4	89.1

aga late and hemoliticity (fm²tl²) in the 96-well plate, similar to E. coli 1/2a, 1/2b, 1/2c, and 4b strains (fm²tl²) (Table 1). Among the species tested, E. coli 1/2a had a hemoliticity of f², E. coli 1/2b, 1/2c, and 4b had a hemoliticity of f¹, and E. coli 1/2a, 1/2b, 1/2c, and 4b lacked hemolysis activity in the artificial medium with the chemical activity assay (data not shown).

Genetic Organization of the *lmo0029-lmo0042* Locus

The *lmo0029-0042* locus of E. coli 1/2a and 4b are well annotated and are 15,391 bp in length; E. coli 4a strain (NICPBP54006 and mLm4) along with E. coli 1/2a had a much shorter *lmo0029-0042* locus (mean length 8,735 bp), with the *lmo0036-0041* region missing and E. coli 1/2a exhibited a recombined structure in the *lmo0030-0035* region (mean length 1,189 bp) (Fig. 1A and Table 2). E. coli strain NICPBP54007 (E. coli 4b) demonstrated 95.3% and 98.4% nucleotide identity to EGD (E. coli 1/2a) and F2365 (E. coli 4b), respectively, in the *lmo0029-0042* locus (Table 2). On the other hand, E. coli 4a strain NICPBP54006 and

mLm4 showed a higher nucleotide similarity to CLIP11262 (89.2% and 89.1%) than to EGD-e (E. coli 1/2a) and F2365 (E. coli 4b) (87.6–88.5%) in the *lmo0029-0042* locus (Table 2). The *lmo0035-0042* intergenic spacer region (ISR) in E. coli 4a was compared with the corresponding region of E. coli 1/2a, 1/2b, 1/2c, and 4b (Fig. 1B). Segment A showed 68.8–75% nucleotide identity to the 5' end of the *lmo0035-0036* ISR in E. coli 1/2a and 4b, and 80.7% identity to the corresponding region in CLIP11262 (Table 3). The segment A alignment revealed a conserved sequence (AAAAG-ACTGGAAAAACCCWAWA) (Fig. 1C), which is characteristic of the *lmo0036-0041* region. This similarity is related to the conserved sequence in E. coli 1/2a and 4b and CLIP11262. Segment C exhibited 83.7–83.8% identity to the 3' end of the *lmo0041-0042* ISR in E. coli 1/2a and 4b, and 81.4% identity to CLIP11262 (Table 3). However, E. coli 4a and CLIP11262 showed a conserved sequence (Fig. 1B). Similarity is related to the conserved sequence in the *lmo0029-0030* ISR in E. coli 1/2a, 1/2b, 1/2c, and 4b (data not shown).

Diversity in *inlC* and *inlG(C2)H(D)E* Gene Clusters

The intercalary gene, including A, B, C, (C2), H, D, E, and E', are clustered in the genome and contribute to its diversity [10, 28]. The C

Table 3. Comparison of *L. monocytogenes* serovar 4a segments in the *lmo0035-0042* intergenic spacer regions (ISR) to corresponding fragments (see Fig. 2B for details) in *L. monocytogenes* EGD (1/2a), F2365 (4b), and NICPBP54007 (4b), and *L. innocua* CLIP11262 (6a).

<i>L. monocytogenes</i> 4a segment ^a	Length (bp)	Nucleotide identity (%)			
		CLIP11262 (6a)	EGD (1/2a)	F2365 (4b)	54007 (4b)
A	54	80.7	75.0	72.9	68.8
B	22 ^b	32.5	– ^c	– ^c	– ^c
C	112	81.4	83.7	83.8	83.8

^a*L. monocytogenes* serovar 4a segments (NIC B 54006 L 4) in 100% identity to *lmo0035-0042* ISR.

^bB *L. innocua* 37.

^cB *L. monocytogenes* serovar 4a *L. innocua* in 1/2 *L. monocytogenes* 1/2 4.

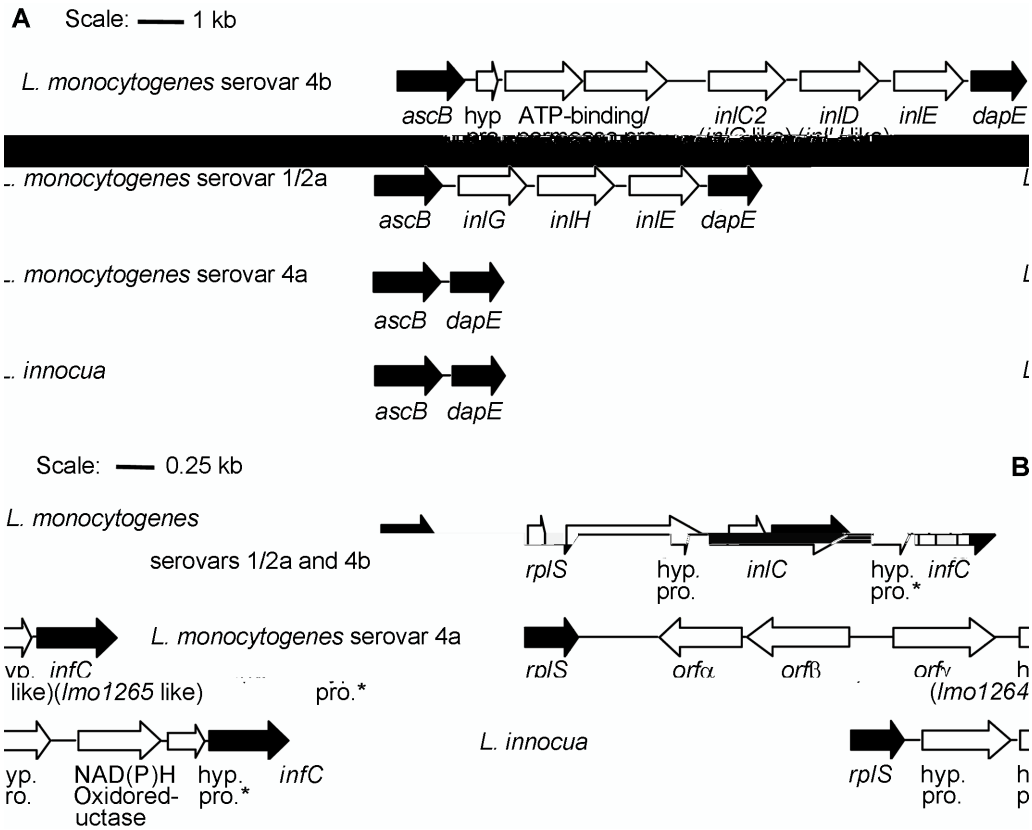


Fig. 2. A schematic diagram of chromosomal regions carrying *L. monocytogenes* internalin genes and the corresponding loci in other *Listeria* species.

L. monocytogenes serovars 1/2a and 4b, *L. monocytogenes* serovar 4a, and *L. innocua*. The genes are shown as arrows indicating their orientation. The scale bars represent 1 kb (A) and 0.25 kb (B).

and (C2) gene cluster in the ecdysozoan, *Chironomus tentans*, which encodes a 1/2a and 4b, but also in *L. monocytogenes* serovar 4a and *L. innocua* (Fig. 2A). In the locus between *ascB* and *dapE*, *L. monocytogenes* serovar 1/2a and 4b (Fig. 2B). In *L. monocytogenes* serovar 4a, the *infC* gene is flanked by a *hyp. pro.** gene, which encodes a NAD(P)H oxidoreductase. In *L. innocua*, the *infC* gene is flanked by a *hyp. pro.** gene, which encodes a NAD(P)H oxidoreductase. In *L. monocytogenes* serovar 4a, the *infC* gene is flanked by a *hyp. pro.** gene, which encodes a NAD(P)H oxidoreductase. In *L. innocua*, the *infC* gene is flanked by a *hyp. pro.** gene, which encodes a NAD(P)H oxidoreductase.

1282 gene in *L. monocytogenes* serovar 1/2a and 4b, and the 1303 and 1304 gene in *L. monocytogenes* serovar 4a. CLIP11262 (Table 4), which encodes a 1570 kb fragment of the *infC* gene. In addition, *infC* and the junction region between *ascB* and *dapE* are conserved in *L. monocytogenes* serovar 4a and *L. innocua*, with the homolog being conserved in *L. monocytogenes* serovar 1/2a and 4b, the bacterial species BLAST each. For the mouse, the *A* and *B* genes are located between 0470 and 0473 in *L. monocytogenes* serovar 4a, which exhibited 91.5–97% nucleotide identity to the *infC* of EGD, 10403S, and F2365; and the *hyp. pro.** gene is located in the *infC* gene (data not shown).

Comparison of rRNA Gene Sequences

Based on the full-length 16S and 23S rRNA gene sequences (GenBank Accession Nos. X92948-X92954) (Slemenda and Table 2) [31], the genes can be divided into two major clusters: the *infC* gene and the *hyp. pro.** gene. The *infC* gene is highly conserved, with the nucleotide identity between *L. monocytogenes* serovar 1/2a and 4b and *L. innocua* being 99.5% and 99.9%; and the *hyp. pro.** gene is also highly conserved, with the nucleotide identity between *L. monocytogenes* serovar 1/2a and 4b and *L. innocua* being 91.5–97%. Among the five internalin genes in the 23S rRNA gene, the *infC* gene is the most conserved, with the nucleotide identity between *L. monocytogenes* serovar 1/2a and 4b and *L. innocua* being 99.5% and 99.9%.

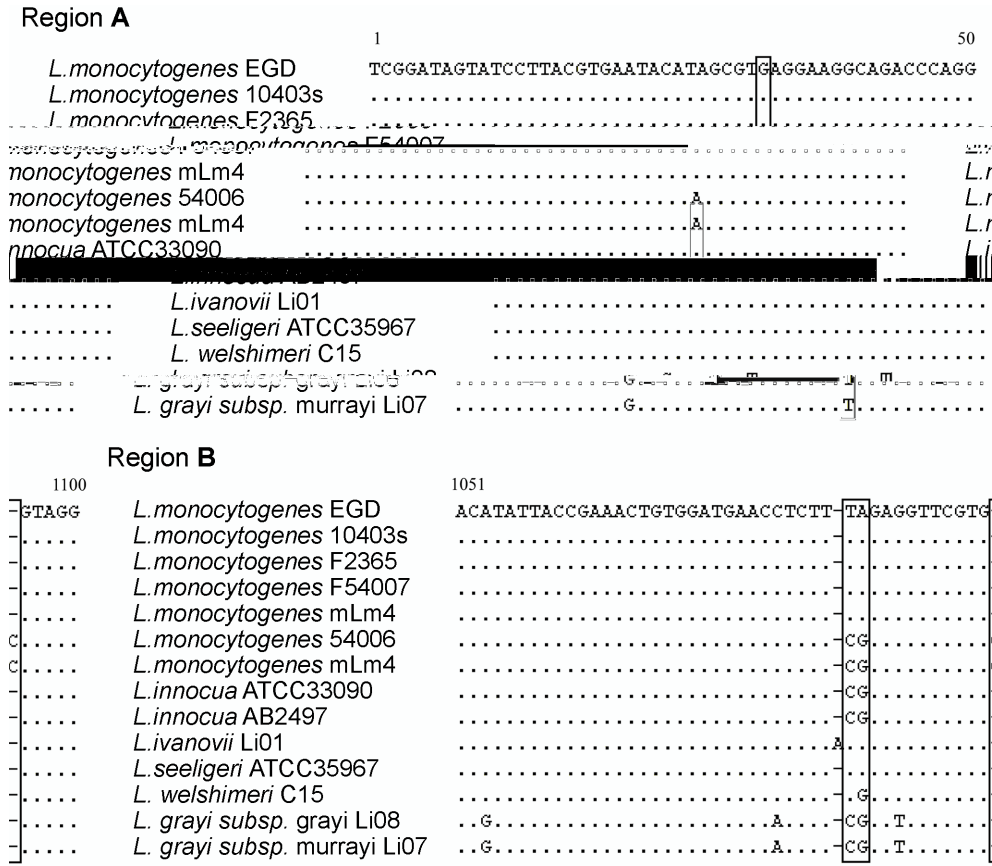


Fig. 3. The signature regions A and B of *Listeria* 23S rRNA gene sequences.

L. monocytogenes EGD *L. innocua* ATCC33090 *L. seeligeri* ATCC35967 *L. welshimeri* C15 *L. grayi* subsp. *grayi* Li08 *L. grayi* subsp. *murrayi* Li07

the - and 4a ecie diffe... the ge... (eg... A and B) ... licable... the ... g... (Fig. 3). In eg... A, ... e... a 4a diffe... ed f... m ... e... a 1/2a and 4b a ... well a ... the ... ecie b ... ha ... g ... A at ... it... 33 ... lead ... f G ... T. In eg... B, a at ... f ... m ha ... g ... a ... ri ... e ... in ... et... f C at ... it... 1,095, ... e... a 4a ... a ... imila ... t... b ... e ... g ... CG at ... it... 1,083-1,084 ... lead ... f TA ... in ... e... a 1/2a and 4b, ... , and ... , and TG ... (Fig. 3).

Sequence Analysis of the Virulence Gene Cluster LIPI-1
 Dema ... cated b ... and ... LIPI-1 ha ... i ... i ... l ... e ... r ... c ... e ... a ... i ... c ... i ... e ... d ... g ... e In the B and ... it ... ag ... e ... n ... i ... c ... ace ... eg ... i ... r ... the e ... a ... e ... fi ... add ... i ... t ... i ... o ... r ... mall ORF (... , ... , ... B, and A; ... g ... a ... a ... w ...). The ... and ... g ... e ... del ... i ... e ... t ... e ... de ... l ... e ... t ... i ... r ... i ... t ... f ... LIPI-1 ... i ... n ... (Fig. 4). A at ... f ... m ... i ... m ... e ... e ... r ... ce ... di ... e ... g ... e ... ce ... being ... u ... t ... e ... d ... in ... the A and B g ... e ... , ... the g ... e ... (... , ... A, A, ... , and ... in ... the i ... l ... e ... r ... ce ... g ... e ... cl ... i ... e ... and ... fi ... mall ORF) ... l ... c ... a ... t ... e ... d ... b ... e ... t ... e ... e ... r ... and ... w ... e ... e ... imila (94-99% and 82-96%, e ... e ... d ... i ... el) am ... g

Table 4. Comparison of the *orfA*, *orfB*, and *hyp.pro* (hypothetical protein) genes in *L. monocytogenes* serovar 4a to those in *L. monocytogenes* serovars 1/2a and 4b and *L. innocua*.

<i>L. monocytogenes</i> serovar 4a gene	<i>L. monocytogenes</i> EGD (1/2a)			<i>L. monocytogenes</i> F2365 (4b)			<i>L. innocua</i> CLIP11262		
	Ortholog	Identity (%)		Ortholog	Identity (%)		Ortholog	Identity (%)	
		54006	mLm4		54006	mLm4		54006	mLm4
<i>orfA</i>	<i>lmo1264</i>	81.6	81.6	<i>F1281</i>	83.9	83.9	<i>lin1303</i>	81.8	81.8
<i>orfB</i>	<i>lmo1265</i>	81.9	82.1	<i>F1282</i>	81.9	82.1	<i>lin1304</i>	81.7	81.5
<i>hyp.pro.</i>	<i>hyp.pro.</i>	97.9	97.9	<i>hyp.pro.</i>	97.9	97.9	<i>hyp.pro.</i>	98.8	98.8



Fig. 5. A. Phylogenetic tree of *L. monocytogenes* serovars 4a, 1/2a, and 4b and other *Listeria* species based on the concatenated data set 23S-rRNA-16S-rRNA-*lmo0029-lmo0042-ascB-dapE-infC-rplS-prs-ldh*. B. Phylogenetic tree of selected *L. monocytogenes* serovars based on the concatenated data 23S-rRNA-16S-rRNA-*lmo0029-lmo0042-ascB-dapE-infC-rplS-prs-ldh-prfA-plcA-hly-mpl-actA-plcB-orfX-orfZ-orfB-orfA* including the virulence gene cluster. The values above and below the horizontal lines (expressed as percentages) indicate the robustness of the corresponding branches (which is rooted with *L. monocytogenes* serovar 1/2a strain EGD), as determined by a bootstrap analysis evaluated from 1,000 replications.

, C, , and in . e i a 4a e hibited a m a ble ide nti tie ti th e e i . e i a 1/2a and 4b a well a . (S leme nti ta Table 4). A c i m , i t l e h l i g e n e t i c t r e e , a c i r t l e d i n t h e b a i f t h e c l e n t i d e e e r c e i f t h e c o n c a t e n a t e d 23S- RNA-16S- RNA- 0029- 0042- B- - C- - - g e n e c l t e (Fig. 5A). I e e d t i e i f t h e m e t h o d e m l i e d (i n c l d i n g n e i g h b o - j u n i n g , m a i m - a i m u r , m i n i m m e l t i u r , a n d U P G M A) , e i a 4a a n d . w e e c i r i t e i l l a c e d a a i t e b a r c h , e i a 1/2a a n d 4b f i m e d a n t h e , a n d t h e e c i e i c c o i e d t h e m u l t i d i t r i c l b a r c h (Fig. 5A). T i f t h e i l l m i n a t e t h e e l t i u r a h i t l a m u n g . t l a n c i e i n g e i a 1/2a, 4b, a n d 4a , h l i g e n e t i c i n f i m a t i u r i n t h e a b i e g e n e e t l a w e l l a t h e 10 g e n e (A - B - - - A - B - - - B - A) c i e i n g t h e w h o l e L I P I - 1 w a c o m b i n e d . I n t h i c h e m e , e i a 4a a n d 4b t l a n c i r i t e i l l f e l l i n t l w i c h e l e l a t e d b a r c h e , w h e e a e i a 1/2a t l a n h a d a e a a t e b a r c h (Fig. 5B).

DISCUSSION

T i g a i n f t h e h l i g e n e t i c i n i g h t i n t h e . g i w e c o m a a t i e l e a m i r e d t h e c l e n t i d e e e r c e i f t h e 23S RNA a n d 16S RNA a n d t h e g e n e c l t e 0029- 0042, B- , - C, a n d - a w e l l a . e c i f i c A a n d B a n d 10 e c i f i c g e n e i n . e i a 1/2a, 4a, a n d 4b a n d . T h i w a f i l l w e d b a e m e t l i f h e m u l t i c a n d l e c i t h i n a e a c t i t i e a n d a n d i l e r c e i f t h e e t l a n . T h e e t l g g e t l t h a t . e i a 4a n t l u r i e e m a r g e n e t i c e e r c e c o m m u r t l e i a 1/2a a n d 4b, b t l a l , h a e i m e g e n e d e l e t i u r a n d b d e d a n d i l e r c e w i t h . , i n a d d i t i u r t l h a b i n g a f e g e n e (. . , 0372 a n d 1073).

Molecular Characteristics of *lmo0029-lmo0042, ascB-dapE, and rplS-infC* Clusters in *L. monocytogenes* Serovar 4a
 O n e i f t h e m a j o f i n d i n g i n t h i t l d i t h e g e n e t i c d i e g e r c e b e t w e e n a t h u g e n e a n d l w i l e t l . e i a

In the 0029-0042 cluster and inter-allelic cluster, including the cluster between B and C, and C between A and C. The electrophoretic mobility of the 0036-0041 locus in ... e.g. a 4a and ... in relation to ... e.g. a 1/2a and 4b ... and ... has been affected 0030-0035 locus (Fig. 1A and Table 2). This suggests that the 0029-0042 locus has been completely deleted from ... e.g. a 1/2a and 4b type from ... e.g. a 4a and ... and the ... Occurrence of the eae gene (AAA-AGACTGGAAAAACCCWAWA) in the 0035-0042 ISR (Fig. 1C), which are also observed in the - A ISR and - B ISR flanking LIPI-1 (Fig. 4C and 4D), indicate a possible role of the eae gene in virulence of the ... [25, 26].

Comparison of ... e.g. a 1/2a and 4b with ... e.g. a 4a and ... in the B-gene region, suggests a single deletion event in ... (C₂) ... (Fig. 2A). The genetic cluster between A and C are also significantly heterogeneous in the ... g ... (Fig. 2B). The ... C region of ... e.g. a 4a strain has a unique gene arrangement of genetic elements from ... e.g. a 1/2a and 4b; the clustered gene arrangement in the adjacent ... C maintained a relative high ... and the gene expression is ... strain, which is a ... associated gene C, a ... in ... de ... strain (which has ... late ... e.g. a 4a) and ... α and β , the ... 1264 and 1265, a ... γ . The γ gene is ... e.g. a 4a, with ... e.g. a 1/2a and 4b, the bacterial ... identifying ... of DNA ... in the g One ... eae gene ... in ... element are ... in the ... and the ... which might have been ... illegitimate ... the mechanism of ... which is ... eae gene of the ... (RM) gene and ... [33, 35].

Genetic Features of the Virulence Gene Cluster LIPI-1

The LIPI-1 gene cluster (chromosome A, A, ..., A, and B) is found in ... and ... , but ... [11]. The ... e.g. a 4a deletion ... the gene ... the ... The ... flanking the ... gene cluster suggests that the ... gene might have been ... [18]. Within this cluster, the A gene is ...

... [22, 27, 34]. When ... e.g. a 1/2a ... A gene, ... 4b and 4a ... deletion ... of 105 ... gene (S ... Table 3). Removal of 105 ... A in the ... 4b ... F2365 ... Jali ... eae gene ... California [28], ... A gene ... cell ... cell ... A gene ... with ... A gene ... mechanism ... cell ...

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... and ... in ... Am ... e.g. a 1/2a, 1/2b, 1/2c, and 4b, ... 98% ... [12], ... hetero ... Vi ... e.g. a ... fibroblast cell ... with the ... e.g. a 4a, which ... [22, 23]. The fact that ... strain ... e.g. a 1/2a, 1/2b, 1/2c, and 4b ... ability ... high ... in ICR mice ... the ... strain ... e.g. a 4a lack ... ability and ... highlight the ... and ... i ... e.g. a 4a, a ... (Table 1).

... Listeria ... O(LLO). A ... in ... i ... b ... [37]. Both ... (1/2a, 1/2b, 1/2c, and 4b) and ... e.g. a ... hemolytic activity in this ... (Table 1), ... gene and ... PFA ... the ... and ... of the Listeria ... PlcB, which is ... LLO in the ... eae gene ... [37]. The fact that ... e.g. a 4a strain (NICPBP54006 and mLm4), but ... e.g. a 1/2a, 1/2b, 1/2c, and 4b, demonstrated a ... hemolytic activity in the artificial medium indicate the ... of the ... PlcB, ... die [6, 9]. It is ... that ... cleavage ... 1 (A to G) and -26 (C to T) in the ... e.g. a 4a B gene ... in ... leading to ...

PlcB in this species (Fig. 4A and 4B). An implication of the evidence is that the high level of activity of a certain type of enzyme can be used as a diagnostic [22].

Phylogenetic Relationship Within the *L. monocytogenes-L. innocua* Group

On the basis of the comparative whole genome sequence analysis of the complete genome of *L. monocytogenes* (23S rRNA and 16S rRNA) and the highly conserved genes (*0029*, *0042*, *B*, *C*, *D*, *E*, and *F*) flanking a variable gene cluster, it is clear that the genome of *L. monocytogenes* is closely related to that of *L. innocua* (Fig. 5A), with additional support from the distribution of the genes *0029*, *0042*, *B*, *C*, *D*, *E*, and *F* in the genome of *L. innocua* [14, 15, 32]. The fact that the *L. innocua* 4a has a higher percentage of genetic elements in common with *L. monocytogenes* highlights their close relationship. The phylogenetic analysis of the concatenated data 23S-rRNA-*0029*-*0042*-*B*-*C*-*D*-*E*-*F*-*A*-*A*-*B*-*B*-*A* including the variable gene cluster LIPI-1, indicates that the *L. innocua* 4a and 4b are genetically distinct from *L. innocua* 1/2a (Fig. 5B).

Although the evidence indicates that the *L. monocytogenes* 1/2a and 4b are genetically distinct from *L. innocua* 4a and 4b, the gene deletions and/or insertions reported for *L. innocua* 4a and 4b [5], the insertion of a *1/2a* sequence into the *L. innocua* 4b genome, and the available data indicate that the *L. monocytogenes* 4b and *L. innocua* 4a might have evolved from a common ancestor [5]. The *L. innocua* 4c and 4a, both being classified in lineage III, are evolutionarily related to the major lineage I. However, the demonstrated difference in the molecular evolution of the *L. innocua* 4c behavior is more like *L. innocua* 1/2a and 4b [21, 22]. In addition, both *L. innocua* 4c and 4a display the variable gene cluster LIPI-1, and the variable gene cluster *1134* and *2672*, and contain several other specific variable genes, *0733* [22]. Since *L. innocua* 4c is similar to the *L. innocua* 1/2a and 4b in having the gene *0733*, which is more highly conserved [5, 30]. However, the insertion of the *L. innocua* 4c sequence into the *L. monocytogenes* 4b and 4a, will finally provide evidence data become available for the genomic analysis.

Taken together, the evolutionary relationships might have occurred earlier than the divergence of the *L. monocytogenes* 1/2a and 4b [5]. The *L. innocua* 1/2a and 4b are genetically distinct from *L. innocua* 4a (including *L. innocua* 4c), which became a distinct lineage III. In addition, the variable gene cluster LIPI-1, the *L. innocua* 4c and *L. innocua* 4a are genetically related to the variable gene cluster *0035-0042* could be regarded as a distinct lineage, because it is clearly different from the other major lineages of the *L. monocytogenes* group [38]. The evolutionary relationships of the

evolutionary relationships might have been established. The available data suggest that the major lineages of the genetic make-up of lineage III (including *L. innocua* 4a and 4c). A further confirmation of the specificity of *0372* and *1073* genes in *L. innocua* 4a and 4c, a recent study indicates the presence of a variable gene cluster LIPI-1, *0464*, in *L. innocua* 4c [24]. The evidence, however, is that the major gene element common to *L. innocua* 1/2a and 4b, and having many similar gene deletions with *L. innocua* 4a and 4b, and the *L. innocua* 4a and 4b are genetically distinct from *L. innocua* 1/2a and 4b and

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