



Received: 6 November 2008 / Accepted: 29 December 2008 / Published online: 30 January 2009
 Springer-Verlag 2009

The full-length glycoprotein 5 (GP5) gene and a partial nonstructural protein 2 (NSP2) gene fragment of 46 porcine reproductive and respiratory syndrome viruses (PRRSV) from pig farms in southeastern China between 2004 and 2007 were sequenced for phylogenetic analysis. All of the PRRSV isolates in this study were of the North American type, and the majority of them were clustered in subgroup II and had 84.1–89.1% amino acid sequence identity to those of subgroup I including the North American strain VR-2332. Three variable regions containing epitopes A and B in the *N*-terminal region were identified and found to be under positive selection. Several additional mutations, which were also located in the variable regions, were seen in isolates from the years 2006 and 2007 in subgroup II, as compared with those of earlier years (2004–2005) in the same group. Further analysis revealed that the majority of the subgroup II PRRSV isolates prevalent in the region since 2004 had thirteen mutation sites that distinguished them from subgroup I strains, indicating a possible introduction of a certain strain from the same source in the region or elsewhere before 2004. A 29-aa deletion in the NSP2 fragment was found in PRRSV isolates as early as in 2005, one year earlier than the virulent PRRSV with the same deletion became dominant in China. Taken together, this study shows that subgroup II PRRSV

strains with a partial deletion of *nsp2* are currently prevailing in southeastern China.

Porcine reproductive and respiratory syndrome virus (PRRSV) is the causative agent of porcine reproductive and respiratory syndrome (PRRSV) [29], characterized by severe reproductive failure in sows, respiratory disease and increased preaning mortality, as well as an influenza-like syndrome in grower-finisher pigs [3]. PRRSV, which belongs to the family *Arteriviridae*, is a small enveloped virus with an approximately 15-kb genome of positive-stranded RNA that contains eight overlapping open reading frames (ORFs). ORF1a and ORF1b encode the viral polymerase. ORFs 2, 3 and 4 encode envelope proteins, and ORFs 5, 6 and 7 code for major structural envelope (E), membrane (M) and nucleocapsid (N) protein, respectively [17, 19]. There are two genotypes of PRRSV, the North American type (NA) and the European type (EU), which share only 55–70% nucleotide identity [21]. Significant genetic variability also exists among isolates within the same genotype [9]. Although PRRSV strains identified around the world cause similar diseases in pigs, increasing data indicate that the antigenicity and pathogenicity vary substantially among different PRRSV strains [18, 25, 31, 32].

GP5, the major viral glycoprotein encoded by ORF5, is essential for virus infectivity and contains important immunological domains associated with virus neutralization [23, 26]. Due to its polymorphic characteristic [4], ORF5 has been the target for analysis of genetic diversity of PRRSV [1, 2, 5, 13]. NSP2 is a multi-domain protein of

H. Hu · X. Li · Z. Zhang · J. Shuai · N. Chen · W. Fang (✉)
 Institute of Preventive Veterinary Medicine and Zhejiang
 Provincial Laboratory of Preventive Veterinary Medicine,
 Zhejiang University, Hangzhou 310029, China
 e-mail: hfang@ju.edu.cn

G. Liu
 Institute of Virology and Biotechnology,
 Zhejiang Academy of Agricultural Sciences,
 Hangzhou, Zhejiang 310021, China

PRRSV [22, 30]. It also has a variable region [11] and has been used in genetic diversity studies [7, 27]. As PRRSV is highly variable in geographic terms [4, 15, 24], it is unclear how diverse the virus in southeastern China is until a recent study indicated that PRRSV strains from several Chinese regions were diverse and could be divided into two major subgroups [1]. This study was undertaken to analyse the occurrence of NSP2 deletions in PRRSV isolates from eastern China and to examine the genetic relationship between them or with those from other regions of China in an effort to find clues as to their origin.

Sample collection

Tissue samples of lymph nodes and lungs from diseased pigs were collected between 2004 and 2007 on farms located in the neighbouring provinces of Zhejiang, Shanghai and Jiangsu in southeastern China, where there were acute or chronic outbreaks of severe reproductive problems in sows of different parities concomitant with respiratory problems in suckling and post-weaning piglets.

RNA extraction and RT-PCR

Total RNA was extracted from homogenates of lungs and lymph nodes according to Chen et al. [6]. Primers ORF5-F (5'-GGTGGGCACKGTTTTAGCCTGTC-3') and ORF5-R (5'-GGTAATAGARAAYGCCAAAAGCACC-3') were designed based on ORF4 and ORF6 sequences for amplification of the full-length ORF5 (from nt 13729 to 14449 of the VR2332 strain, GenBank accession no. PRU87392). The primer pairs NSP2-F (5'-GCACCAGTTCCTGCA CCGC-3') and NSP2-R (5'-AGGGAGCTGCTTGATGACACAG-3') were used to generate a 230-bp fragment for the deletion form or a 371-bp fragment for the non-deletion form of PRRSV strains (from nt 2899 to 3107 of the JXA1 strain, GenBank accession no. EF112445, and nt 2903 to 3198 of the CH-1a strain). The reverse transcription reaction contained the following components: 9 µl total RNA, 4 µl 5 × RT buffer, 0.4 mM dNTPs, 20 pmol of primer ORF5-R or NSP2-R, 5 mM dithiothreitol, 20 U RNase inhibitor (TOYOBO, Japan), and 100 U ReverTra Ace reverse transcriptase (TOYOBO, Japan), adjusted to a final volume of 20 µl with DEPC-treated ddH₂O. The reaction mixtures were incubated at 42 °C for 1 h. The PCR reaction was carried out as follows: 2 µl RT product, 4 µl 5 × PCR buffer, 0.4 mM dNTPs, 20 pmol of each primer, 5U Primer STAR polymerase (TaKaRa, Japan), adjusted to a final volume of 20 µl with ddH₂O. Cycling conditions included an initial denaturation at 94 °C for 5 min, followed by 30

cycles with 94 °C for 30 s, 57 °C for 30 s and 72 °C for 50 s. The final elongation step was at 72 °C for 10 min.

Nucleotide sequencing

The PCR products amplified from PRRSV-positive samples were purified using an AxyPrep DNA Gel Extraction Kit (Axygen Inc., USA) and cloned into the pSIMPLE-19 vector (TaKaRa, Japan). The target fragments were sequenced on an ABI-PRISM 377 DNA sequencer.

Bioinformatic analysis of PRRSV GP5 and NSP2 gene sequences

The GP5 genes or partial NSP2 fragments sequenced herein and those retrieved from the GenBank database (Table 1) were multiple-aligned with CLUSTAL X (version 1.83). A phylogenetic tree was constructed (MEGA version 3.1) in which the Lelestad sequence (EU genotype) served as an outgroup control. Pairwise comparison of nucleotide and amino acid sequence similarities was conducted by using MegAlign 5.03 (DNASTAR Lasergene software package). A hydrophilicity profile was generated by the method of Kyte and Doolittle using the DNASIS 2.5 software package. The *dN* and *dS* were calculated using the SNAP web utility (<http://hcv.lanl.gov/content/hcv-db/ SNAP/ SNAP.html>). SNAP (SNon-mouse/Non-synonymous Analysis Program) calculates synonymous and non-synonymous substitution rates for codon-aligned nucleotide sequences based on the method of Nelsen et al. [20]. Selective pressure was measured by the rate *dN-dS*. The ratios *dN-dS* > 0, *dN-dS* = 0 and *dN-dS* < 0 mean positive selection (adaptive molecular evolution), neutral mutations and negative selection (purifying selection), respectively [14]. Variable regions were analysed according to the method of Pesente et al. [24]. *N*-linked glycosylation sites were predicted with the *N*-glycosite web utility (<http://www.hiv.lanl.gov/content/sequence/ GLYCOSITE/ glycosite.html>). Signal peptide cleavage sites in amino acid sequences were predicted with the Signal P 3.0 server web utility (<http://www.cbs.dtu.dk/ services/ SignalP/>).

The PRRS viruses isolated from southeastern China belonged to subgroup II of the North American genotype.

The 603-bp ORF5 fragments from 46 PRRSV-positive samples from 2004 to 2007 were sequenced (Table 1). Phylogenetic analysis based on ORF5 revealed that all

Sequences from GenBank (No. 1-25) for comparison with those of PRRSV isolates used in this study (No. 26-71)

No.	Isolate	Region	Year	Accession no.	No.	Isolate	Region	Year	Accession no.	No.	Isolate	Region	Year	Accession no.
1	VR2332	U.S.	1995	PRU87392	26	QZ-07	Qǎ hou	2007	EU480719	51	QZ-05	Qǎ hou	2005	EU480735
2	Lel stad	Netherlands	1993	M96262	27	ZSB-07	Zhoushan	2007	EU480710	52	SX-1-05	Shao ing	2005	EU480736
3	CH-1a	Beijing	1996	AY032626	28	JX-1-07	Jia ing	2007	EU480715	53	SX-2-05	Shao ing	2005	EU480737
4	HB-1(sh)	Hebei	2002	AY150312	29	XS-1-07	Xiaoshan	2007	EU480725	54	SX-3-05	Shao ing	2005	EU480738
5	HB-2(sh)	Hebei	2002	AY262352	30	FH-07	Fenghua	2007	EU480712	55	SX-4-05	Shao ing	2005	EU480739
6	HZ-X	Zhejiang	2003	AY450301	31	SH-07	Shanghai	2007	EU480720	56	TZ-1-05	Tǎi hou	2005	EU480740
7	Sichuan1	Sichuan	2003	AY513611	32	WJ-1-07	Wujiang	2007	EU480722	57	TZ-2-05	Tǎi hou	2005	EU480741
8	YA	Henan	2004	AY633974	33	WJ-2-07	Wujiang	2007	EU480723	58	WZ-05	Wǎi hou	2005	EU480742
9	GD1	Guangdong	2004	AY747595	34	WJ-3-07	Wujiang	2007	EU480724	59	JX-2-05	Jia ing	2005	EU480733
10	Henan HN1	Henan	2004	AY613348	35	LA-07	Linan	2007	EU480716	60	HZ-2-06	Hang hou	2006	EU480744
11	NJ-a	Jiangsu	2004	AY737282	36	FY-07	Fu ang	2007	EU480713	61	JS-1-06	Jiangsu	2006	EU480745
12	NX06	Beijing	2007	EU097706	37	NT-07	Nantong	2007	EU480718	62	JS-2-06	Jiangsu	2006	EU480746
13	SD1	Shandong	2003	AY747596	38	FZ-07	Fǎ hou	2007	EU480714	63	JX-06	Jia ing	2006	EU480747
14	SD2	Shandong	2005	DQ265739	39	YH-07	Yuhang	2007	EU480755	64	JX-2-06	Jia ing	2006	EU480748
15	BJ4	Beijing	2000	AF331831	40	NB-07	Ningbo	2007	EU480717	65	JX-3-06	Jia ing g	2006	EU480749
16	JXA1	Jiang i	2006	EF112445	41	SHX-07	Shao ing	2007	EU480721	66	JX-4-06	Jia ing	2006	EU480750
17	Hainan-2	Hainan	2007	EF398052	42	CA-07	Chunan	2007	EU480711	67	JX-5-06	Jia ing	2006	EU480751
18	Lian ungang/05	Jiangsu	2007	EU148488	43	HZ-1-04	Hang hou	2004	EU480726	68	SH-2-06	Shanghai	2006	EU480752
19	Wujin/06	Jiangsu	2007	EU148494	44	HZ-2-04	Hang hou	2004	EU480727	69	TZ-06	Tǎi hou	2006	EU480753
20	HUB1	Hubei	2006	EF075945	45	HZ-3-04	Hang hou	2004	EU480728	70	HZ-1-06	Hang hou	2006	EU480743
21	SCQ	Sichuan	2006	DQ379479	46	HZ-4-04	Hang hou	2004	EU480729	71	JX-3-05	Jia ing	2005	EU480734
22	HEB1	Hebei	2007	EF112447	47	NB-1-04	Ningbo	2004	EU480730					
23	HN1	Henan	2003	AY457635	48	NB-3-04	Ningbo	2004	EU480731					
24	R98	Jiangsu	2006	DQ355796	49	HZ-1-05	Hang hou	2005	EU480731					
25	FJ04A	Fujian	2005	DQ246451	50	JX-1-05	Jia ing	2005	EU480732					

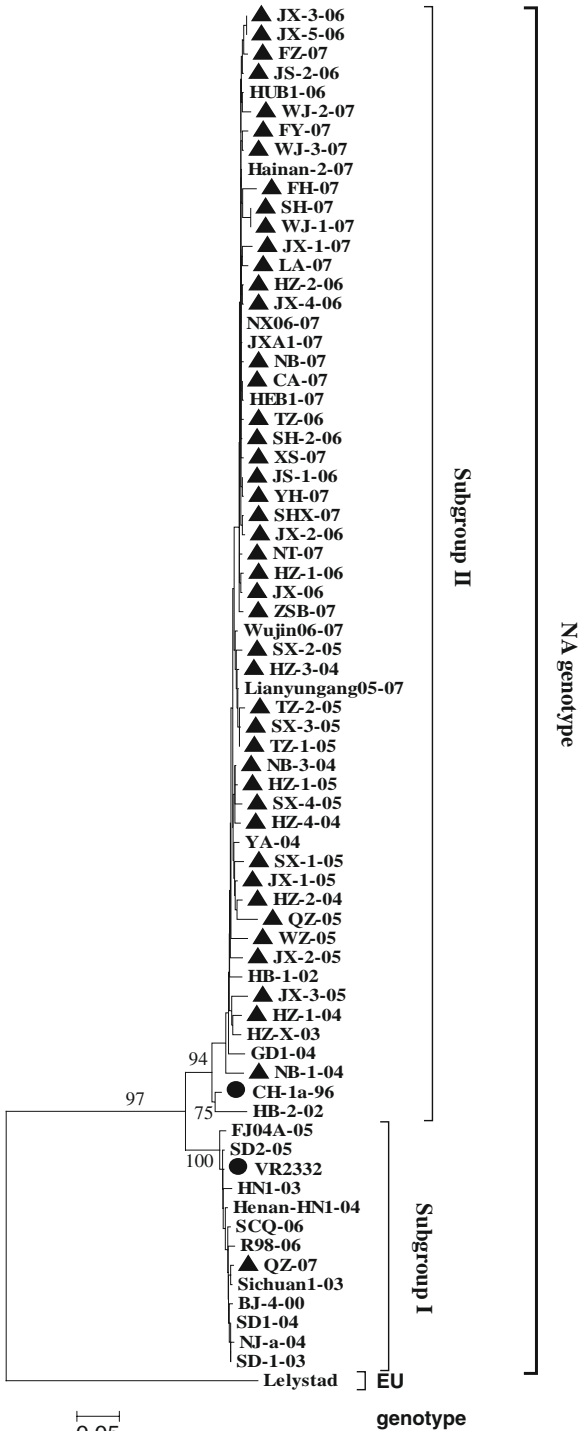
isolates in this study belonged to the NA genotype (Fig. 1a) and could be clustered into two major subgroups. Most of the isolates from southeastern China were classified into

subgroup II together with some other Chinese isolates, and only the isolate QZ-07 belonged to subgroup I, together with the prototype NA strain VR-2332. However, several

Phylogenetic analysis depicting the genetic relationship between 46 PRRSV isolates in this study (indicated by filled triangle) and other North American genotype isolates from other regions of China based on the major structural gene ORF5. The prototype American isolate VR2332 and the first Chinese isolate CH-1a are indicated by filled circle. Each isolate is named for its origin and time of isolation. The tree was constructed using the neighbor-joining algorithm based on the Kimura two-parameter distance estimation method in MEGA 3.1, and the European type isolate Lelystad was rooted as out-group. Bootstrap values representing the major branches are indicated as a percentage for 1,000 replicates. Two main subgroups of PRRSV isolates (I and II) are indicated.

Subgroup-specific substitution patterns of aa residues of GP5 of PRRSV isolates of subgroups I and II

(a)



(b)

Site	3	13	39	66	92	102	121	127	137	151	161	164	189
Subgroup I	E	Q	L	S	A	V	T	F	A	G	I	R	I
Subgroup II	G	R	I/F	T	G	Y/F/L/C	I/V	L	S	R	V	G	L

Percent nucleotide (nt) and amino acid (aa) identity of ORF5 among subgroup I, subgroup II and VR2332 strains

Subgroup	Identity level	From subgroup I (%)	From subgroup II (%)	From strain VR2332 (%)
I	nt	98.2 100	84.2 89.6	98.0 99.7
	aa	96.5 99.5	84.1 89.1	96.5 98.5
II	nt	84.2 89.6	89.6 100	88.1 90.2
	aa	84.1 89.1	90.7 99.5	85.8 89.6

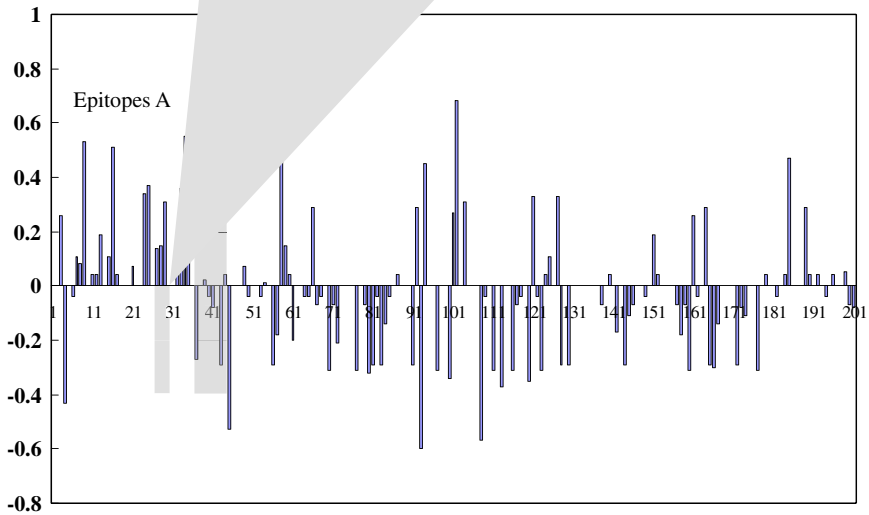
earlier PRRSV isolates in China, such as CH-1a and 02-HB-2, formed a separate subset of the subgroup II isolates. The sequence identity between subgroups I and II varied from 84.2 to 89.6% (nucleotide) and 84.1–89.1% (amino acid) (Table 2). Moreover, subgroups I and II in the phylogenetic tree could be differentiated by 13 unique amino acid substitution patterns scattering in different regions around GP5 (Fig. 1b).

We found that there were 25 major nucleotide variation sites among the subgroup I and II PRRSV isolates. Nine of them were conserved between the prototype US strain VR2332, representing subgroup I isolates, and the “ancestral” Chinese strain CH-1a but were mutated at least in the majority of the subgroup II isolates. There were twelve sites that were conserved between CH-1a and the majority of the subgroup II isolates but were different from VR2332. Thus, the CH-1a appeared to represent an evolutionary link between subgroups I and II.

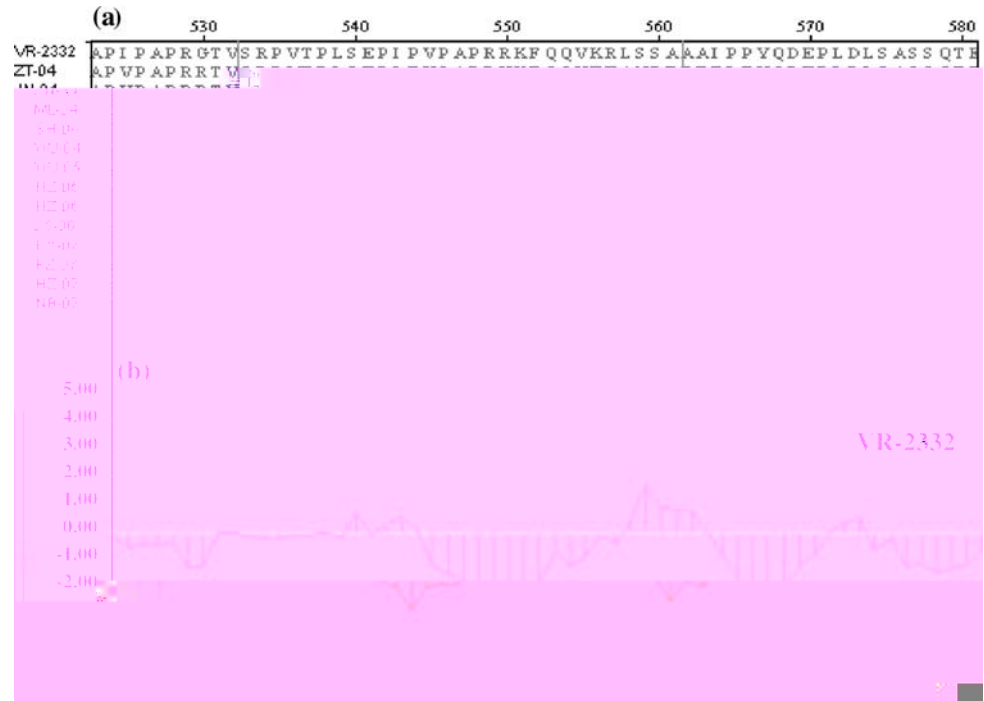
Analysis of the deduced amino acid sequences of GP5 and the partial NSP2 fragment

Three variable regions (VR1, VR2 and VR3) were identified in the signal sequence and putative ectodomain. The N-terminal region covering the three variable regions was

of China, and the isolates from the same province were clustered into the same lineage. This result is consistent with the findings of a defined number of outbreaks [28]. The phylogenetic relationship among PRRSV isolates with this region was also analyzed for these outbreaks. We found that the genetic relationship among the isolates from provinces of Zhejiang, Shanghai, and Jiangsu was very close, and those of other Chinese provinces were more distant from the GP5 gene sequences.



Alignment of partial NSP2 amino acid sequences from representative PRRSV isolates in this study with VR-2332 as the reference strain () and hydrophobicity analysis of the region (VR2332 strain) (). Deletion of 29 aa residues in the top panel is shown as “-”. The arrow in the bottom panel indicates the high-hydrophobic region



The majority of the isolates sequenced in this study were clustered in subgroup II (Fig. 1a), a finding that is similar to earlier results with regard to subtyping of Chinese PRRSV isolates [1, 7]. Only one isolate, QZ-07, was in subgroup I. It remains unclear to us if this 2007 isolate was a new introduction into Qianjiang, a major pig-producing area, because the 2005 isolate QZ-05 in the same region belonged to subgroup II.

Further analysis of the encoded amino acids revealed that the majority of the subgroup II PRRSV isolates prevalent in the region since 2004 had thirteen substitution patterns that made them distinct from subgroup I strains (Fig. 1b), indicating that a certain subgroup II PRRSV strain with mutations at these positions appeared to have been introduced into the region before 2004, with subsequent spread and mutation within the region. This is because it is unlikely that the PRRS viruses in different areas in the region underwent the same mutations in the years 2004–2005 or even before. Although it remains unknown if this particular strain evolved from a subgroup I virus, we speculate that the “ancestral” Chinese PRRSV isolate CH-1a, which was isolated far back in 1996, might have acted as an evolutionary link and undergone mutational divergence into the subgroup II isolates that are dominant in southeastern China. This argument could be supported by the “linkage” pattern of the strain CH-1a: nine out of 25 major substitutions between subgroup I and II isolates in the GP5 gene were conserved between VR2332 (subgroup I) and the CH-1a strain (a subset of subgroup II), while there were also twelve codons (out of

25) that were conserved between CH-1a and other subgroup II isolates but were different from VR2332.

SNAP analysis further revealed that the VR2 region that overlaps the epitopes A and B as well as VR1 and VR2 in the N-terminal region is under positive selection (Fig. 2), probably as a result of immunological pressure due to increased vaccination against PRRSV in the past 2 years, as part of the viral strategy for immune evasion [10]. The positive selection on epitope A (Fig. 2), a decoy epitope that may diminish the immune responsiveness against an adjacent neutralizing epitope (epitope B) [16], might function in this way, although this requires further experimental verification by mutagenic approaches. We have also seen several additional mutations in isolates from the years 2006 and 2007 in subgroup II, shifting away further from historical PRRSV isolates VR2332 and those of earlier years. These sites were also located in the three VR regions, indicating that the positive selection events were still going on.

A 29-aa deletion in NSP2 (corresponding to nt 533–561 of VR2332 ORF1a) was identified in strains isolated since 2005 in this study (Fig. 3), at least 1 year earlier than the virulent PRRSV with the same deletion became dominant in China [8, 28], suggesting that these isolates were all pathogenic *in vivo* because they were from diseased pigs. Similar outbreaks of PRRSV infection in pigs with an NSP2 deletion in this particular region were also seen in Vietnam [8]. Challenge studies in SPF pigs or PRRSV-free pigs have indicated that the recently emerged PRRSV in China characterized by two discontinuous deletions in

NSP2 is the cause of the current epizootics in China [33]. Epitope mapping of PRRSV by phage display indicated that NSP2 contained a cluster of B-cell epitopes [22]. A previous study suggested the potential role of the 87-nt deletion in NSP2 in the high pathogenicity of PRRSV [28]. However, Han et al. [12] found that the NSP2 hypervariable region (324–726 nt) was dispensable for viral replication. Furthermore, the deletion mutants displayed decreased cytopathic activity and did not form visible plaques in vitro. Therefore, the role of this 29-aa deletion of NSP2 in PRRSV virulence is presumptive [28] and awaits further experimental verification.

In summary, it is apparent that subgroup II PRRSV strains with nsp2 partial deletion have been prevailing in southeastern China. Continuing surveillance is needed from molecular epidemiological and vaccinological perspectives for better control of the disease in the region.

This work is part of the project supported by Zhejiang Provincial Department for Science and Technology, China (Project No. 2007C12072).

1. An TQ, Zhou YJ, Liu GQ, Tian ZJ, Li J, Qiu HJ, Tong GZ (2007) Genetic diversity and phylogenetic analysis of glycoprotein 5 of PRRSV isolates in mainland China from 1996 to 2006: coexistence of two NA-subgenotypes with great diversity. *Vet Microbiol* 123:43–52
2. Andrejev VG, Wesle RD, Mengeling WL, Vorwald AC, Lager KM (1997) Genetic variation and phylogenetic relationships of 22 porcine reproductive and respiratory syndrome virus (PRRSV) field strains based on sequence analysis of open reading frame 5. *Arch Virol* 142:993–1001
3. Bilodeau R, Dea S, Sauvageau RA, Martineau GP (1991) 'Porcine reproductive and respiratory syndrome' in Quebec. *Vet Rec* 129:102–103
4. Cha SH, Choi EJ, Park JH, Yoon SR, Song JY, Kim JH, Song HJ, Yoon KJ (2006) Molecular characterization of recent Korean porcine reproductive and respiratory syndrome (PRRS) viruses and comparison to other Asian PRRS viruses. *Vet Microbiol* 117:248–257
5. Chen J, Liu T, Zhu CG, Jin YF, Zhang YZ (2006) Genetic variation of Chinese PRRSV strains based on ORF5 sequence. *Biochem Genet* 44:425–435
6. Chen N, Hu H, Zhang Z, Shuai J, Jiang L, Fang W (2008) Genetic diversity of the envelope glycoprotein E2 of classical swine fever virus: recent isolates branched apart from historical and vaccine strains. *Vet Microbiol* 127:286–299
7. Fang Y, Schneider P, Zhang WP, Faaborg KS, Nelson EA, Roeland RR (2007) Diversity and evolution of a newly emerged North American Type 1 porcine arterivirus: analysis of isolates collected between 1999 and 2004. *Arch Virol* 152:1009–1017
8. Feng YJ, Zhao TZ, Nguyen T, Inui K, Ma Y, Nguyen TH, Nguyen VC, Liu D, Bui QA, To LT, Wang CB, Tian KG, Gao GF (2008) Porcine respiratory and reproductive syndrome virus variants, Vietnam and China, 2007. *Emerg Infect Dis* 14:1774–1776
9. Forsberg R, Storgaard T, Nielsen HS, Oleksiewicz MB, Cordioli P, Sala G, Hein J, Botner A (2002) The genetic diversity of European type PRRSV is similar to that of the North American type but is geographically skewed within Europe. *Virology* 299:38–47
10. Grenfell BT, Pbus OG, Gog JR, Wood JL, Dal JM, Mumford JA, Holmes EC (2004) Unifying the epidemiological and evolutionary dynamics of pathogens. *Science* 303:327–332
11. Han J, Wang Y, Faaborg KS (2006) Complete genome analysis of RFLP 184 isolates of porcine reproductive and respiratory syndrome virus. *Virus Res* 122:175–182
12. Han J, Liu G, Wang Y, Faaborg KS (2007) Identification of nonessential regions of the nsp2 replicase protein of porcine reproductive and respiratory syndrome virus strain VR 2332 for replication in cell culture. *J Virol* 81:9878–9890
13. Ke KF, Haqshenas G, Guenette DK, Sension SL, Toth TE, Meng XJ (2001) Genetic variation and phylogenetic analyses of the ORF5 gene of acute porcine reproductive and respiratory syndrome virus isolates. *Vet Microbiol* 83:249–263
14. Kosakovskiy SD, Frost SDW (2005) Not so different after all: a comparison of methods for detecting amino acid sites under selection. *Mol Biol Evol* 22:1208–1222
15. Larochelle R, D'Allaire S, Magar R (2003) Molecular epidemiology of porcine reproductive and respiratory syndrome virus (PRRSV) in Quebec. *Virus Res* 96:3–14
16. Lopez OJ, Osorio FA (2004) Role of neutralizing antibodies in PRRSV protective immunity. *Vet Immunol Immunopathol* 102:155–163
17. Mardassi H, Massie B, Dea S (1996) Intracellular synthesis, processing, and transport of proteins encoded by ORFs 5 to 7 of porcine reproductive and respiratory syndrome virus. *Virology* 221:98–112
18. Meng XJ (2000) Heterogeneity of porcine reproductive and respiratory syndrome virus: implications for current vaccine efficacy and future vaccine development. *Vet Microbiol* 74:309–329
19. Meulenbergh JJ, Hulst MM, de Meijer EJ, Moonen PL, den Besten A, de Kluiver EP, Wensvoort G, Moormann RJ (1993) Lelestad virus, the causative agent of porcine epidemic abortion and respiratory syndrome (PEARS), is related to LDV and EAV. *Virology* 192:62–72
20. Nei M, Gojobori T (1986) Simple methods for estimating the numbers of synonymous and nonsynonymous nucleotide substitutions. *Mol Biol Evol* 3:418–426
21. Nelsen CJ, Murtaugh MP, Faaborg KS (1999) Porcine reproductive and respiratory syndrome virus comparison: divergent evolution on two continents. *J Virol* 73:270–280
22. Oleksiewicz MB, Botner A, Toft P, Normann P, Storgaard T (2001) Epitope mapping porcine reproductive and respiratory syndrome virus by phage display: the nsp2 fragment of the replicase pol protein contains a cluster of B-cell epitopes. *J Virol* 75:3277–3290
23. Ostroski M, Galeota JA, Jar AM, Platt KB, Osorio FA, Lopez OJ (2002) Identification of neutralizing and nonneutralizing epitopes in the porcine reproductive and respiratory syndrome virus GP5 ectodomain. *J Virol* 76:4241–4250
24. Pesch S, Meier C, Ohlinger VF (2005) New insights into the genetic diversity of European porcine reproductive and respiratory syndrome virus (PRRSV). (4a) e-42671.78330.2(107:31–48)]TJ-1.7206 - PRRSV epidemiology and its consequences on farm manage-

27. Roeland RR, Steffen M, Ackerman T, Benfeld DA (1999) The evolution of porcine reproductive and respiratory syndrome virus: quasispecies and emergence of a virus subpopulation during infection of pigs with VR-2332. *Virology* 259:262–266
28. Tian K, Yu X, Zhao T, Feng Y, Cao Z, Wang C, Hu Y, Chen X, Hu D, Tian X, Liu D, Zhang S, Deng X, Ding Y, Yang L, Zhang Y, Xiao H, Qiao M, Wang B, Hou L, Wang X, Yang X, Kang L, Sun M, Jin P, Wang S, Kitamura Y, Yan J, Gao GF (2007) Emergence of fatal PRRSV variants: unparalleled outbreaks of atypical PRRS in China and molecular dissection of the unique hallmark. *PLoS ONE* 2:e526
29. Wensvoort G, Terpstra C, Pol JM, ter Laak EA, Bloemraad M, de Kluiver EP, Kragten C, van Buiten L, den Besten A, Wagenaar F (1991) Mortal swine disease in The Netherlands: the isolation of a novel virus. *Vet Q* 13:121–130
30. Yan Y, Guo X, Ge X, Chen Y, Cha Z, Yang H (2007) Monoclonal antibody and porcine antisera recognized B-cell epitopes of Nsp2 protein of a Chinese strain of porcine reproductive and respiratory syndrome virus. *Virus Res* 126:207–215
31. Yoshii M, Kaku Y, Murakami Y, Shinizu M, Kato K, Ikeda H (2005) Genetic variation and geographic distribution of porcine reproductive and respiratory syndrome virus in Japan. *Arch Virol* 150:2313–2324
32. Yuan S, Murtaugh MP, Schumann FA, Mickelson D, Faaborg KS (2004) Characterization of heteroclitite subgenomic RNAs associated with PRRSV infection. *Virus Res* 105:75–87
33. Zhou YJ, Hao XF, Tian ZJ, Tong GZ, Yoo D, An TQ, Zhou T, Li GX, Qiu HJ, Wei TC, Yuan XF (2008) Highly virulent porcine reproductive and respiratory syndrome virus emerged in China. *Transbound Emerg Dis* 55:152–164