



Protective efficacy of a novel recombinant *Platyedon grandiflorum* virus-like particle vaccine against hepatitis B virus infection in mice

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ARTICLE INFO

Article history:

Received 4 September 2008

Received in revised form 31 October 2008

Accepted 3 November 2008

Available online 27 November 2008

Keywords:

Protective efficacy

Platyedon grandiflorum

Adjuvant

Hepatitis B surface antigen (HBsAg)

Cell-mediated immunity

T1/T2 lymphocytes

ABSTRACT

The protective efficacy of a novel recombinant *Platyedon grandiflorum* virus-like particle (VLP) vaccine against hepatitis B virus (HBV) infection in mice was evaluated. The VLP vaccine was composed of the HBV surface antigen (HBsAg) and the *Platyedon grandiflorum* virus-like particle (VLP) core. The VLP vaccine was adjuvanted with aluminum hydroxide (Al(OH)₃) and lipopolymer (LPS). The VLP vaccine was evaluated for its protective efficacy in mice. The results showed that the VLP vaccine was highly effective in protecting mice against HBV infection. The protective efficacy of the VLP vaccine was significantly higher than that of the HBsAg vaccine. The VLP vaccine induced a strong cell-mediated immune response in mice, which was characterized by a high level of interferon-γ (IFN-γ) production and a high level of T1/T2 lymphocyte proliferation. The VLP vaccine also induced a strong humoral immune response in mice, which was characterized by a high level of anti-HBsAg antibody production. The results suggest that the VLP vaccine is a promising candidate for the prevention of HBV infection in humans.

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1. Introduction

Hepatitis B virus (HBV) is a major cause of liver disease and liver cancer in humans. The HBV infection is a global health problem, with an estimated 2 billion people infected worldwide [1]. The HBV infection is characterized by a high level of chronicity, with up to 90% of infected individuals developing chronic hepatitis B (CHB) [2]. The HBV infection is also characterized by a high level of mortality, with up to 25% of infected individuals developing liver cancer [3]. The HBV infection is a major cause of liver disease and liver cancer in humans. The HBV infection is a global health problem, with an estimated 2 billion people infected worldwide [1]. The HBV infection is characterized by a high level of chronicity, with up to 90% of infected individuals developing chronic hepatitis B (CHB) [2]. The HBV infection is also characterized by a high level of mortality, with up to 25% of infected individuals developing liver cancer [3].

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The results suggest that the VLP vaccine is a promising candidate for the prevention of HBV infection in humans. The VLP vaccine induced a strong cell-mediated immune response in mice, which was characterized by a high level of interferon-γ (IFN-γ) production and a high level of T1/T2 lymphocyte proliferation. The VLP vaccine also induced a strong humoral immune response in mice, which was characterized by a high level of anti-HBsAg antibody production. The results suggest that the VLP vaccine is a promising candidate for the prevention of HBV infection in humans.

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e.s 41157.2()1157.2a.d37

Table 1
Sequences of primers used for RT-PCR.

Gene	Primer sequence	Product size (bp)	Accession
GAPDH	5' AAATGGTGAAGGTCGGTGTG 3' 5' TGAAGGGGTCGTTGATGG 3'	108	NM.001001303
IL-2	5' GCACCCACTTCAAGCTCCA 3' 5' AAATTGAAGGTGAGCATCCTG 3'	174	NM.008366
IFN-γ	5' CGGCACAGTCATTGAAAGCCTA 3' 5' GTTGCTGATGGCCTGATTGTC 3'	199	NM.008337

GAPDH, glyceraldehyde 3-phosphate dehydrogenase.

the (ABC). After incubation for 30 min, the absorbance was read at 450 nm using a microplate reader. The reaction was stopped by adding 100 μl of 10% SDS. The absorbance was read at 450 nm.

2.11. Real-time RT-PCR for cytokine gene expression

Sequences of primers used for real-time RT-PCR are as described before. Cells were seeded in 24-well plates at a density of 5×10^6 cells per well. After 24 h of culture, the cells were treated with HBsA (final concentration 4 μg/ml) as added. After 24 h of culture, the cells were harvested and total RNA was extracted using RNeasy spin columns (Qiagen, Crawley, UK). Total RNA (1 μg) was reverse transcribed using Superscript II reverse transcriptase (Invitrogen, Carlsbad, CA, USA) in the presence of 5% CO₂ in a 96-well plate.

99 T s[(99)][(99 (d)sfe)]T.99 a 37

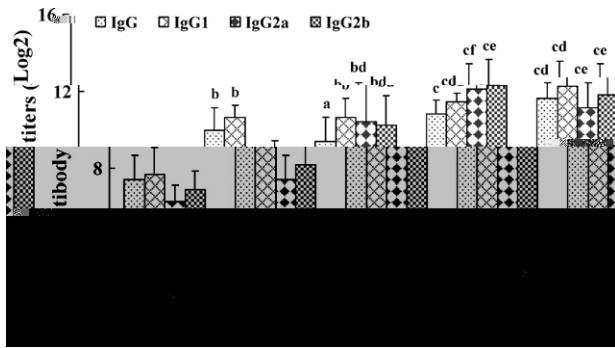


Fig. 3. Effect of a chimeric D (PD) on HBsAg specific IgG, IgG1, IgG2a, and IgG2b antibody titers in mice immunized with HBsAg. Sealed 2 weeks after immunization, mice were sacrificed and sera were collected. The effect of PD on HBsAg specific IgG, IgG1, IgG2a, and IgG2b antibody titers was determined by ELISA as described. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.05$, $^b P<0.01$, and $^c P<0.001$.

IgG, IgG2a, and IgG2b antibody titers were significantly higher ($P<0.001$) in mice immunized with HBsAg/A. than in mice immunized with HBsAg. The effect of PD on HBsAg specific IgG, IgG1, IgG2a, and IgG2b antibody titers was determined by ELISA as described. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.05$, $^b P<0.01$, and $^c P<0.001$. Mice immunized with HBsAg/A. showed a significant increase in antibody titers compared to mice immunized with HBsAg. The effect of PD on HBsAg specific IgG, IgG1, IgG2a, and IgG2b antibody titers was determined by ELISA as described. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.05$, $^b P<0.01$, and $^c P<0.001$.

3.3. Effects of PD on NK cell activity in mice immunized with HBsAg

The effect of PD on NK cell activity in mice immunized with HBsAg was determined by LDH release assay. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.05$, $^b P<0.01$, and $^c P<0.001$.

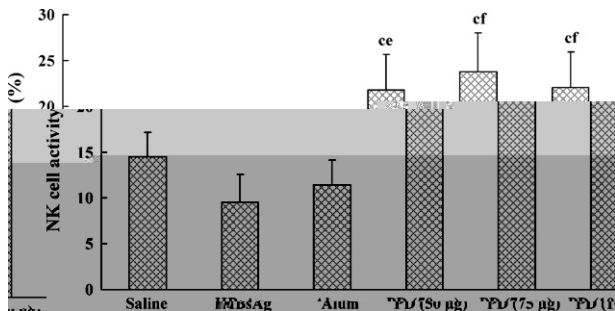


Fig. 4. Effect of a chimeric D (PD) on NK cell activity in mice immunized with HBsAg. Sealed 2 weeks after immunization, mice were sacrificed and spleen cells were collected. The effect of PD on NK cell activity was determined by LDH release assay as described. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.001$, $^b P<0.01$, and $^c P<0.001$.



Fig. 5. Effect of a chimeric D (PD) on CTL activity in mice immunized with HBsAg. Sealed 2 weeks after immunization, mice were sacrificed and spleen cells were collected. The effect of PD on CTL activity was determined by LDH release assay as described. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.001$ and $^b P<0.001$, respectively.

CTL activity was significantly higher ($P<0.001$) in mice immunized with HBsAg/A. than in mice immunized with HBsAg. The effect of PD on CTL activity was determined by LDH release assay as described. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.001$ and $^b P<0.001$, respectively.

3.4. Effects of PD on specific CTL activity in mice immunized with HBsAg

The effect of PD on specific CTL activity in mice immunized with HBsAg was determined by LDH release assay. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.001$ and $^b P<0.001$, respectively. Mice immunized with HBsAg/A. showed a significant increase in specific CTL activity compared to mice immunized with HBsAg. The effect of PD on specific CTL activity was determined by LDH release assay as described. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.001$ and $^b P<0.001$, respectively.

3.5. Effect of PD on cytokine secretion by splenocytes from HBsAg-immunized mice

In order to assess the effect of PD on cytokine secretion by splenocytes from HBsAg-immunized mice, the levels of IL-2, IFN- γ , and IL-10 were determined by ELISA. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.001$, $^b P<0.01$, and $^c P<0.001$. Mice immunized with HBsAg/A. showed a significant increase in cytokine secretion compared to mice immunized with HBsAg. The effect of PD on cytokine secretion was determined by ELISA as described. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.001$, $^b P<0.01$, and $^c P<0.001$.

3.6. Effect of PD on mRNA expression of cytokines in splenocytes from HBsAg-immunized mice

Since PD significantly increased IgG2a and IgG2b antibody titers and CTL activity, the effect of PD on mRNA expression of cytokines in splenocytes from HBsAg-immunized mice was determined by RT-PCR. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.001$, $^b P<0.01$, and $^c P<0.001$. Mice immunized with HBsAg/A. showed a significant increase in cytokine mRNA expression compared to mice immunized with HBsAg. The effect of PD on cytokine mRNA expression was determined by RT-PCR as described. The data are expressed as mean \pm S.E. ($n=5$). Statistical differences between HBsAg and HBsAg/A. were as follows: $^a P<0.001$, $^b P<0.01$, and $^c P<0.001$.

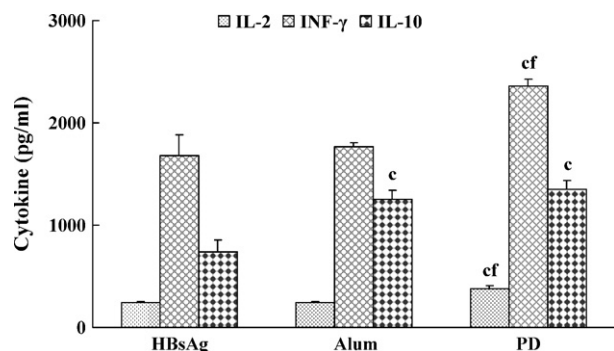


Fig. 6. Effect of adjuvant D(PD) on HBsA and cytokine levels. Mice were immunized with HBsA (4 μ g) and adjuvant (HBsA, Alum, or PD) (4 μ g) on days 0, 14, and 28. Serum samples were collected on day 28 and analyzed for cytokine levels (IL-2, INF- γ , and IL-10) by ELISA. The results are expressed as mean \pm S.E. ($n=5$). Significant differences between HBsA and HBsA/A are indicated as $P<0.001$ and $P<0.001$, respectively.

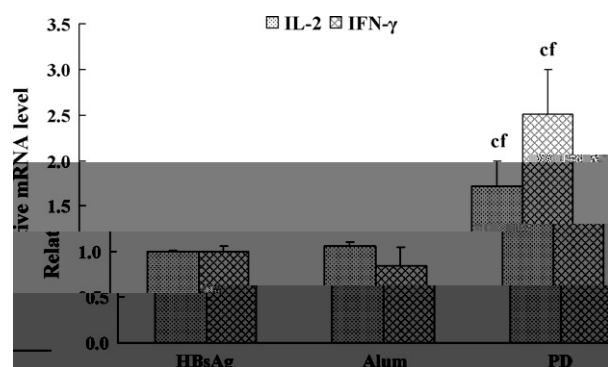


Fig. 7. Effect of adjuvant D(PD) on RNA levels of IL-2 and IFN- γ in HBsA. Mice were immunized with HBsA (4 μ g) and adjuvant (HBsA, Alum, or PD) (4 μ g) on days 0, 14, and 28. Total RNA was extracted on day 28 and analyzed for IL-2 and IFN- γ mRNA levels by RT-PCR. The results are expressed as mean \pm S.E. ($n=5$). Significant differences between HBsA and HBsA/A are indicated as $P<0.001$ and $P<0.001$, respectively.

The results of HBsA and HBsA/A are shown in Table 1 ($P<0.001$). The results of IL-2 and IFN- γ RNA levels are shown in Table 2 ($P<0.05$). The results of HBsA/A and HBsA are shown in Table 3. The results of PDs are shown in Table 4. The results of HBsA/A and HBsA are shown in Table 5.

4. Discussion

The results of the present study show that the T1 and T2 adjuvants, which are composed of HBsA and alum, can be used as adjuvants for HBsA [15,16]. The results of the present study show that the T1 and T2 adjuvants can significantly increase the levels of IL-2, INF- γ , and IL-10 in the serum of mice [17]. The results of the present study show that the T1 and T2 adjuvants can significantly increase the levels of IL-2, INF- γ , and IL-10 in the serum of mice [18]. The results of the present study show that the T1 and T2 adjuvants can significantly increase the levels of IL-2, INF- γ , and IL-10 in the serum of mice [19,20]. The results of the present study show that the T1 and T2 adjuvants can significantly increase the levels of IL-2, INF- γ , and IL-10 in the serum of mice [21]. The results of the present study show that the T1 and T2 adjuvants can significantly increase the levels of IL-2, INF- γ , and IL-10 in the serum of mice [22].

5, IL 10 and IL 13. The results of the present study show that the T1 and T2 adjuvants can significantly increase the levels of IL-2, INF- γ , and IL-10 in the serum of mice [23].

The results of the present study show that the T1 and T2 adjuvants can significantly increase the levels of IL-2, INF- γ , and IL-10 in the serum of mice [24]. The results of the present study show that the T1 and T2 adjuvants can significantly increase the levels of IL-2, INF- γ , and IL-10 in the serum of mice [25]. The results of the present study show that the T1 and T2 adjuvants can significantly increase the levels of IL-2, INF- γ , and IL-10 in the serum of mice [26].

de Δ s a e d a PD Δ d a e d e a Δ f e e s Δ ses,
a d e c e d a b a a c e d T 1/T 2 e e s Δ s e Δ HBsA c e
a s a s s Δ a e d s e s e a e a c e e Δ I G2a, I G2b a d
I G1 e e s [29].

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e e Δ e d e a d a a c Δ PD, e a a e d e
T 1/T 2 c Δ e s e c e Δ f i e s Δ HBsA e d c e
s ELISA. PD Δ s f i c a c e a s e d e d c Δ f
T 2 c Δ es IL 10, b a s Δ s e a c e d e d c Δ
 Δ T 1 c Δ es IL 2 a d IFN γ f Δ s e Δ c e s e HBsA
e d c e. H IL 2 s e c e Δ c e a e d e d c Δ
 Δ a a e s e c f i c c e a Δ f e a e e s Δ se, e e
e e Δ IFN γ s c Δ s s e e c e a s e Δ I G2a a d
I G2b a b d e s. S a e c e e d A a d
PD a d e e Δ IL 10 a d c Δ es Δ d e d e e s
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 Δ f e e c d a e e e c a s e s Δ s b e f e c a e s
e a Δ s Δ T 1 c Δ es, e e d e a e RT PCR

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